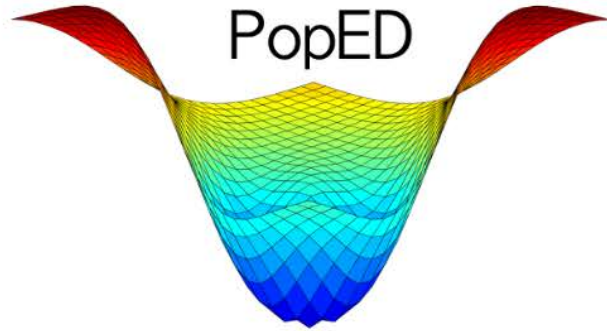


A Gentle Introduction to Optimal Design for Pharmacometric Models



with PopED, PFIM, and mrgsolve

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Why are we here?

- You want to design a study
- You'll be fitting a model to the results
- You don't want that model to fail spectacularly
- You don't have the time or the patience to run a bunch of simulations



Outline

- Optimal design background
- Software tools
- Simple model with PopED and PFIM
 - Evaluation
 - Optimization
 - Simulation
- More complex models with PopED and mrgsolve



Optimal design background



Meet the Fisher information matrix (FIM)

$$M_F(\Psi, \xi) = - \mathbb{E} \left[\frac{\partial^2}{\partial \Psi \partial \Psi^T} \log L(\Psi; y) \middle| \Psi \right]$$

where

- Ψ is the vector of population parameters (e.g., THETAs, OMEGAs, and SIGMAs in NONMEM),
- y is the vector of observations,
- ξ is the vector of design variables (e.g., sampling times), and
- $\log L$ is the log-likelihood.



Fisher in winter coat



Why should I care about that thing?

Cramér-Rao lower bound:

$$\text{cov}(\hat{\Psi}) \geq [M_F(\Psi, \xi)]^{-1}$$

when $\hat{\Psi}$ is an unbiased estimator of Ψ .

- Lower bounds for relative standard errors (RSEs) can be obtained from the diagonals of the inverse of the FIM
- This means we have a quick way of evaluating (lower bounds on) the precision of our parameter estimates.



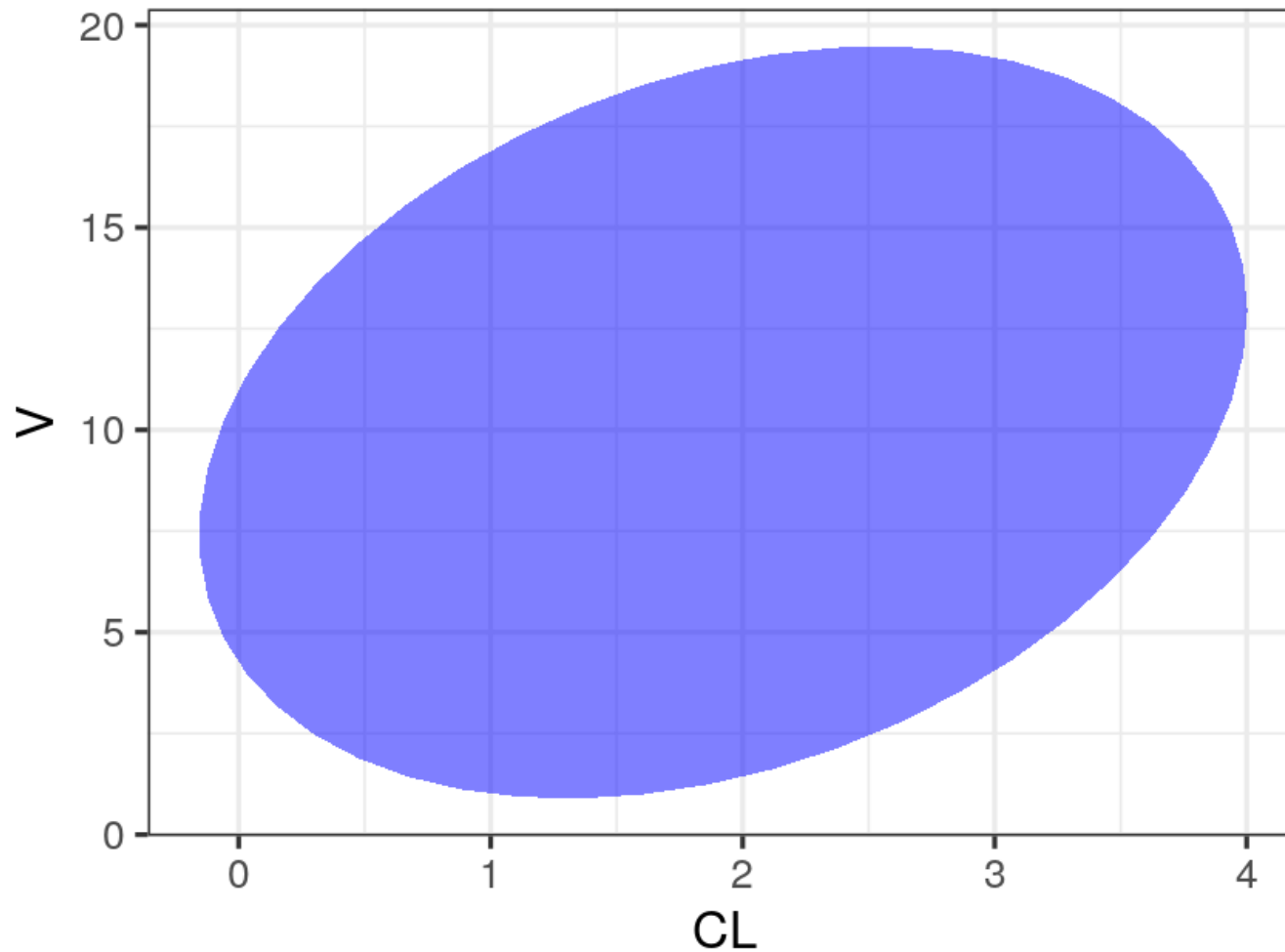
OK, but really. Why should I care about that thing?

$$\text{cov}(\hat{\Psi}) \geq [M_F(\Psi, \xi)]^{-1}$$

- D -optimality criterion
- D -optimal designs maximise the determinant of the FIM
- Equivalent to minimising the volume of the confidence ellipsoid of the parameter estimates
- Huh?



This is a confidence ellipsoid in 2 dimensions



Catch-22 of optimal design

- For linear models, the dependence of $M_F(\Psi, \xi)$ on Ψ disappears
- No such luck for nonlinear models
- In order to design our experiment in a way that will produce the best parameter estimates, we first need to know the values of those parameters



Nonlinear mixed effects models are even more problematic

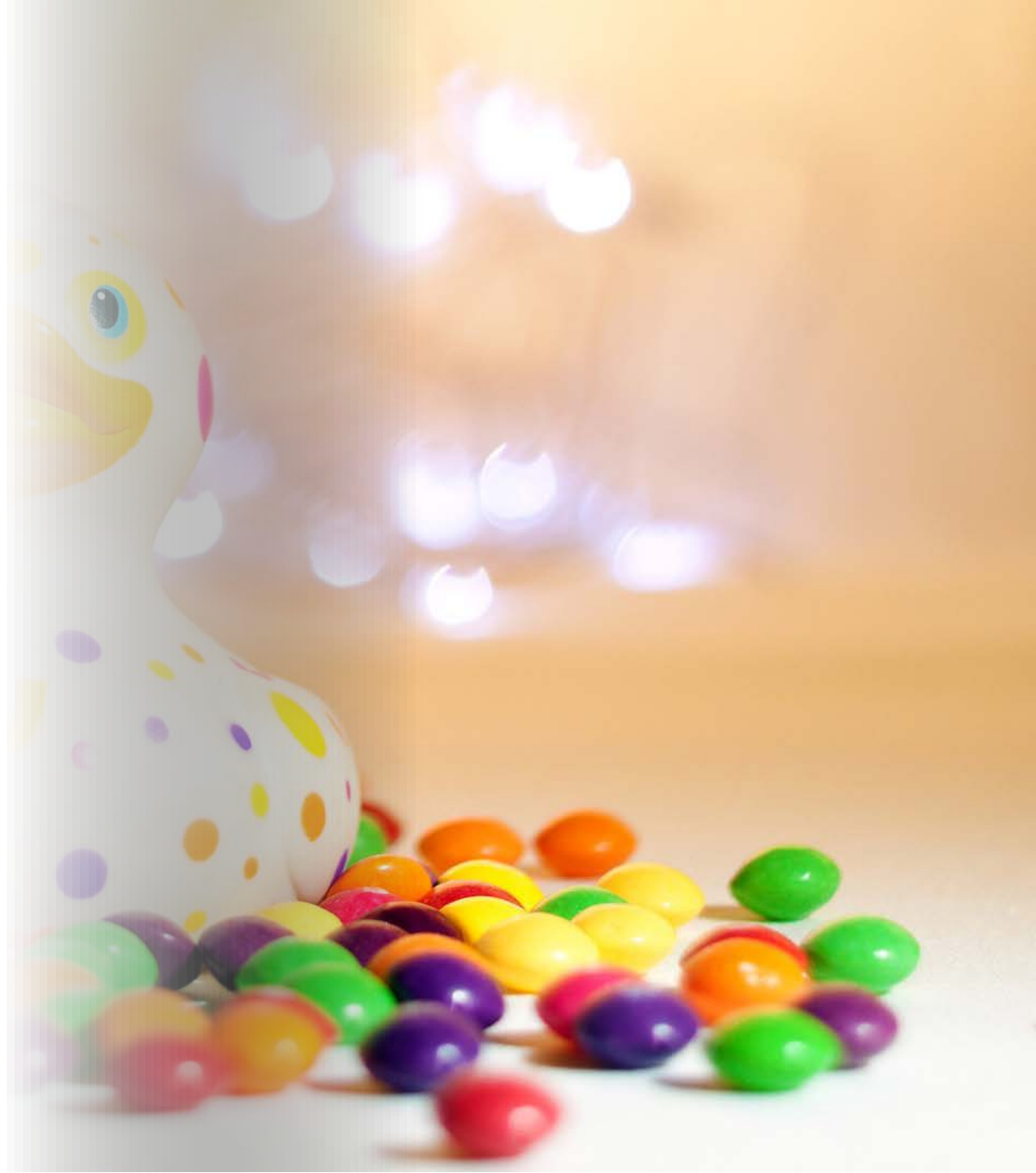
- No analytic expression for the likelihood, so we rely on approximations
- So our FIM is
 - an approximation
 - to a lower bound
 - that depends on the parameter values
- But...
- All is not lost
- Usually we have adequate information on parameter estimates
- Approximate lower bounds are usually not far off from values obtained from simulation
- More to come on simulation...

Mentre, Mallet, and Baccar (1997)
Retout, Duffull, and Mentre (2001)
Retout and Mentre (2003)

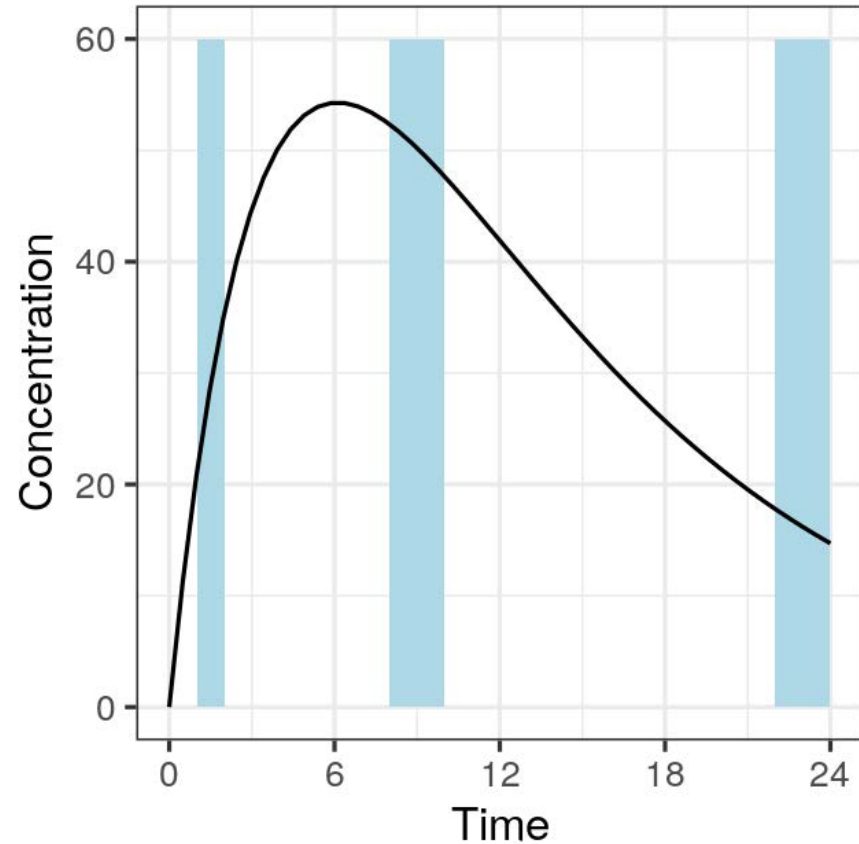


Evaluation vs Optimisation

- Optimal design can be used to optimise a study design (duh)
- We can also just use the FIM to quickly evaluate a design by calculating RSEs
- Optimisation is often a last resort (we can just evaluate a few candidate designs in many situations)
- Sometimes resources are too tightly constrained or our intuition isn't good enough to find feasible designs without optimising using a search algorithm



Sampling windows



- "Optimal" sampling times are often not practical
- Even without optimisation, we can't always collect samples at precise times
- Sampling windows can be optimised or determined manually



Tools for optimal design

Feature	Software				
	PFIM	PkStaMp	PopDes	PopED	POPT
Language	R	Matlab	Matlab	Matlab FreeMat	Matlab FreeMat
Available on website	✓		✓	✓	✓
Library of PKPD models	✓	✓	✓	✓	✓
User-defined models	✓	✓	✓	✓	✓
Multiresponse models	✓	✓	✓	✓	✓
Designs differ across responses	✓	✓	✓	✓	✓
ODE models	✓	✓	✓	✓	✓
Full FIM	✓	✓	✓	✓	–
Full covariance matrix for Ω	–	✓	✓	✓	–
Full covariance matrix for Σ	–	–	✓	✓	–
IOV	✓	–	✓	✓	–
Discrete covariates/power	✓/✓	–	✓/–	✓/✓	✓/–

Abbreviations are as follows: FIM, Fisher information matrix; GUI, graphical user interface; IOV, interoccasion variability; ODE, ordinary differential equation; PKPD, pharmacokinetic–pharmacodynamic; Σ , residual covariance matrix; Ω , interindividual covariance matrix.

Notable exception: NONMEM \$DESIGN

Understanding the Methodology and Uses of Some New Features
in NONMEM 7.5: Clinical Trial Design Evaluation and Optimization,
and Delay Differential Equations

July 14, 2021 @ 11:00 am - 12:00 pm

ISoP MCS SIG Webinar Series

presents

Robert Bauer, Ph.D.

Pharmacometrics and PK/PD Modeling & Simulation, ICON Clinical Research, LLC.

Seminar Title

**Understanding the Methodology and Uses of Some New Features in NONMEM 7.5: Clinical Trial Design
Evaluation and Optimization, and Delay Differential Equations**

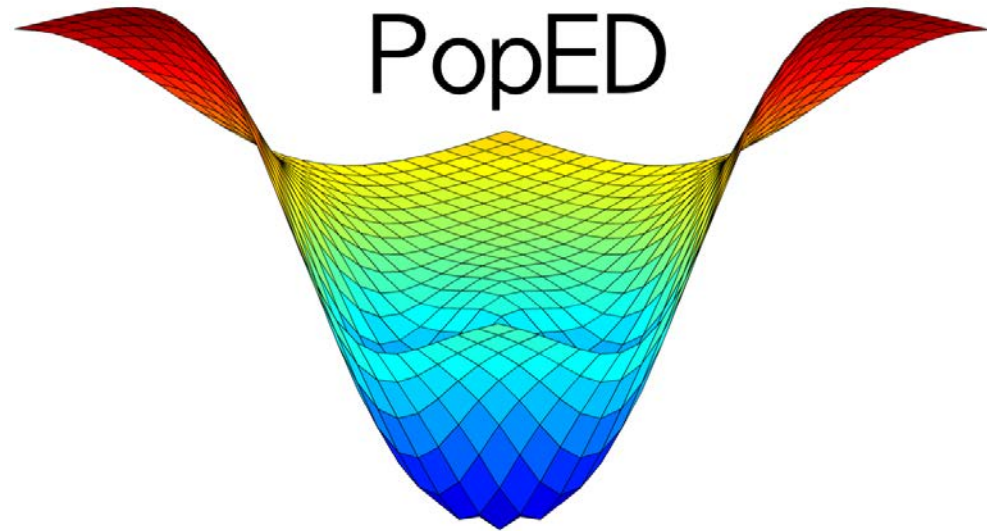
July 14, 2021 @ 11 AM EST

Recording: <https://sites.google.com/view/mcssig/mcs-sig#h.v7bdq6l7d86v>

Bauer, Hooker, and Mentre (2021)



PopED



- <https://andrewhooker.github.io/PopED/>
- Originally in O-Matrix and Matlab, now an R package

Foracchia, Hooker, Vicini, and Ruggeri (2004)
Nyberg, Ueckert, Strömberg, Hennig, Karlsson, and Hooker (2012)



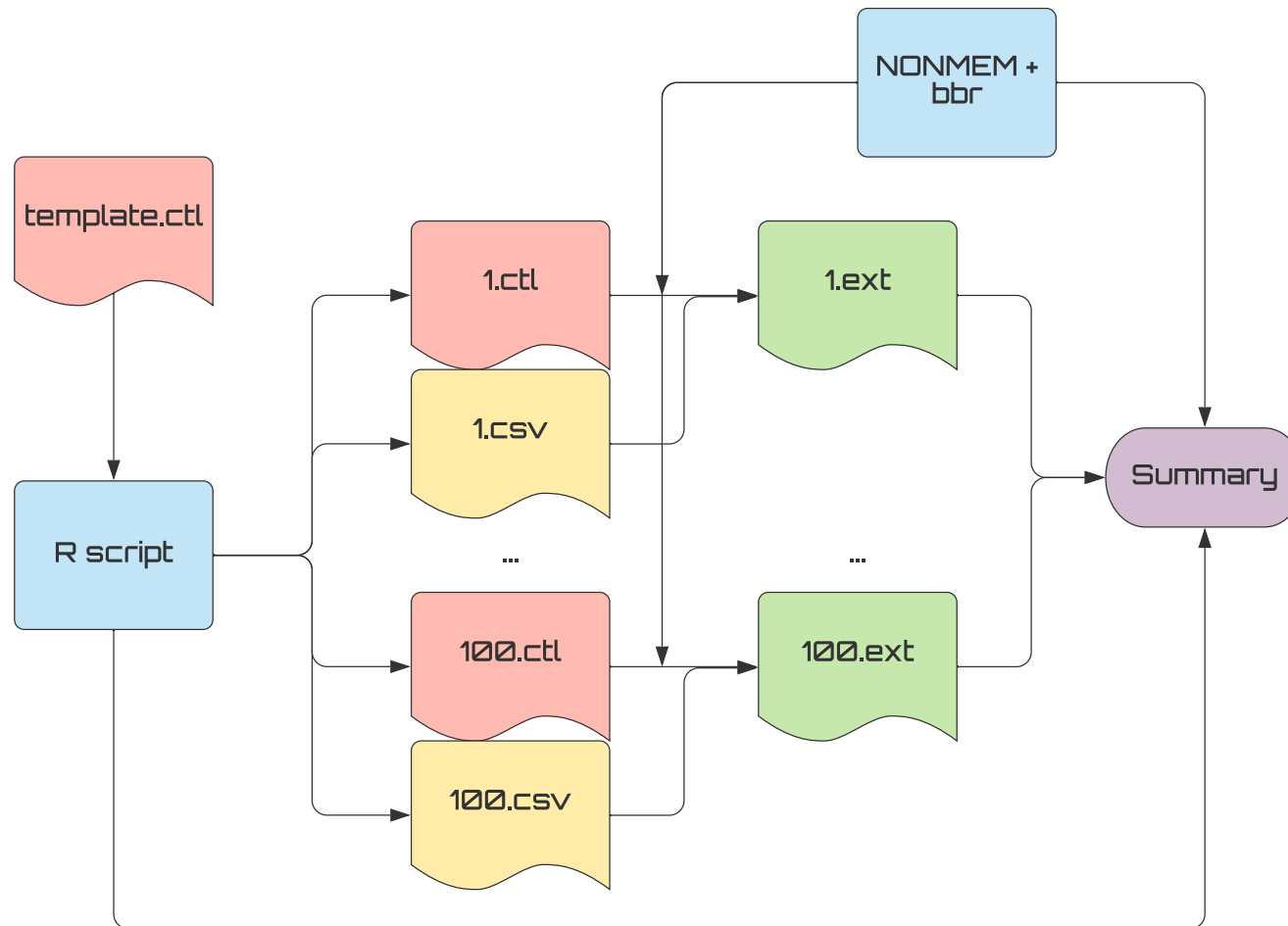
PFIM



- <http://www.pfim.biostat.fr/>
- Previously set of R functions, now an R package (Version 5.0)
- Version 4.0 also available as GUI: PFIM Interface



SSE: Stochastic Simulation and Estimation



Example: Closed-form PK model



Introducing our example

Mockdrozaline has been studied in adult subjects, and we now must design a study in pediatric patients.

A study objective is to evaluate the PK in this new population, but PK sampling is necessarily sparse.

Our mission is to ensure that these samples are timed such that we can sufficiently estimate the PK parameters in pediatric patients.

- Population
 - 12 subjects
 - Aged 6 to < 12
 - Expected median weight of 32 kg
- Treatment
 - 10 mg QD mockdrozaline for 24 weeks
- PK samples
 - Proposed samples:
 - 5 hours postdose on Day 1;
 - predose on Weeks 8, 12, 24; and
 - 168 hours after the final dose



The model

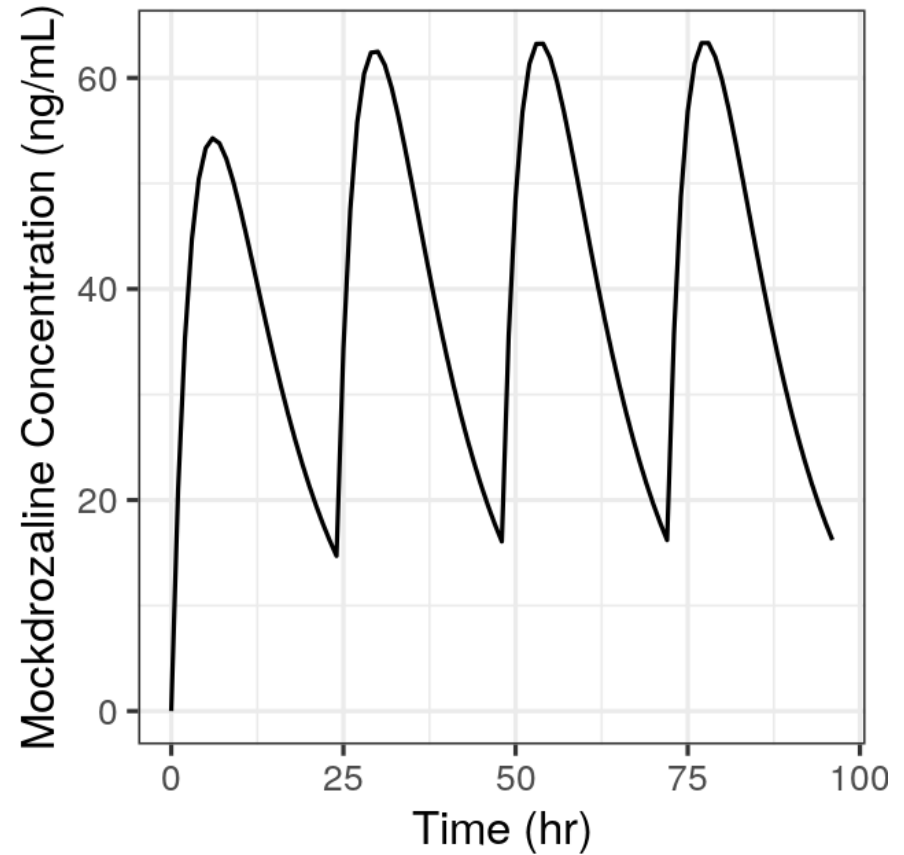
1-compartment model with 1st-order absorption and weight covariates on CL and V:

CL	V	KA	wt_cl	wt_v
10	100	0.25	0.75	1

Log-normal IIV on CL, V, and KA; additive & proportional residual error:

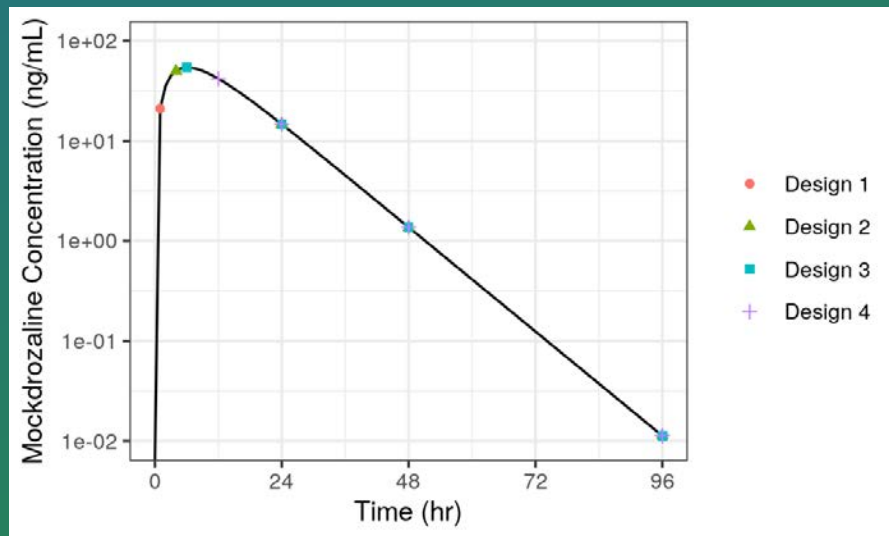
om_CL	om_V	om_KA	sigma_prop	sigma_add
0.08	0.1	0.2	0.05	1

(these are variances)



Pop Quiz

Which of these designs will give us the best* RSE for KA, assuming a single 10 mg dose in 10 adult (70 kg) subjects?



1. 1, 24, 48, 96 hours

2. 4, 24, 48, 96 hours

3. 6, 24, 48, 96 hours

4. 12, 24, 48, 96 hours

[*]According to FIM
 $t_{\max} \approx 6$ hours



Pop Quiz

Which of these designs will give us the best* RSE for KA, assuming a single 10 mg dose in 10 adult (70 kg) subjects?

- Design 1 (first sample at 1 hours): RSE = 23.4%
- Design 2 (first sample at 4 hours): RSE = 27.1%
- Design 3 (first sample at 6 hours): RSE = 30.5%
- Design 4 (first sample at 12 hours): RSE = 48.9%

[*]According to PopED



The PopED setup

PopED requires 3 functions in order to define a model:

- `ff()`, the structural model;
- `fg()`, the parameter model (including IIV and IOV);
- `feps()`, the residual error model.

`create.poped.database()` collects together these model functions, parameter values, and everything related to study design.



PopED: Plot of initial design

```
plot_model_prediction(  
  poped_db,  
  model.names = c("Day 1", "Steady state"),  
  facet_scales = "free_x",  
  model_num_points = 200  
) +  
  labs(x = "Time from dose (h)") +  
  theme_bw()
```



PopED: Evaluate FIM

```
FIM <- evaluate.fim(poped_db)
det(FIM)
```

```
. [1] 0.04804071
```

```
get_rse(FIM, poped_db)
```

```
.           CL           V           KA           d_CL           d_V           d_KA  SIGMA[1,1]  S
. 2.983332e+05 3.132099e+06 4.936376e+06 5.192188e+01 4.888612e+02 6.485818e+02 2.997297e+01 4.0
```



PopED: Tweak timepoint and evaluate FIM

```
poped_db2 <- create.poped.database( poped_db, xt = c(5, c(23, 24, 24, 168)) )  
FIM2 <- evaluate.fim(poped_db2)  
get_rse(FIM2, poped_db2)
```

.	CL	V	KA	d_CL	d_V	d_KA	SIGMA[1,1]	SIGMA[2,2]
.	15.48778	142.08530	223.35925	50.88485	418.53695	549.75962	29.68444	40.82396



PopED: D -optimal design: Starting from the original design

```
poped_db <- create.poped.database(  
  ...  
  xt = c(5, c(rep(24, 3), 168)),  
  minxt = c(0, c(rep(23, 3), 96)),  
  maxxt = c(6, c(rep(24, 3), 168)),  
  ...  
)  
output <- poped_optim(  
  poped_db,  
  opt_xt = TRUE,  
  parallel = TRUE,  
  parallel_type = "multicore",  
  seed = 1  
)  
summary(output)
```

```
. =====  
. FINAL RESULTS  
. Optimized Sampling Schedule  
. Group 1: Model 1: 0.3275  
. Group 1: Model 2:      23      24      24      96  
.   
. OFV = 20.2291  
.   
. Efficiency:  
. ((exp(ofv_final) / exp(ofv_init))^(1/n_parameters)) = 18  
.   
. Expected relative standard error  
. (%RSE, rounded to nearest integer):  
.   Parameter   Values      RSE_0   RSE  
.           CL       10    298333   38  
.           V       100   3132099  149  
.           KA      0.25   4936376  153  
.          d_CL     0.08      52    50  
.          d_V     0.1     489   204  
.          d_KA     0.2     649   127  
.   SIGMA[1,1]    0.05      30    30  
.   SIGMA[2,2]      1     41    41  
.   
. Total running time: 18.044 seconds
```



PopED: Add sample after final (SS) dose

```
poped_db_final <- create.poped.database(  
  poped_db_extra_ss,  
  xt = c(5, c(rep(24, 3), 72, 168)),  
  minxt = c(0, c(rep(23, 3), 0, 168)),  
  maxxt = c(6, c(rep(24, 3), 168, 168))  
)
```

```
FIM_final <- evaluate.fim(poped_db_final)  
get_rse(FIM_final, poped_db_final)
```

.	CL	V	KA	d_CL	d_V	d_KA	SIGMA[1,1]	SIGMA[2,2]
.	12.31062	86.75053	136.64068	50.42738	317.03881	419.93344	29.55819	28.95827



PopED: D -optimal design: Add sample after final (SS) dose

```
output_final <- poped_optim(  
  poped_db_final,  
  opt_xt = TRUE,  
  parallel = TRUE,  
  parallel_type = "multicore",  
  seed = 1  
)  
summary(output_final)
```

```
. =====  
. FINAL RESULTS  
. Optimized Sampling Schedule  
. Group 1: Model 1: 0.4455  
. Group 1: Model 2:      23      23      23  41.09      168  
.   
. OFV = 27.717  
.   
. Efficiency:  
. ((exp(ofv_final) / exp(ofv_init))^(1/n_parameters)) = 2.  
.   
. Expected relative standard error  
. (%RSE, rounded to nearest integer):  
.   Parameter   Values   RSE_0   RSE  
.           CL        10       12     9  
.           V        100       87    14  
.           KA        0.25      137    17  
.          d_CL        0.08       50    48  
.           d_V        0.1      317    74  
.          d_KA        0.2      420    63  
.   SIGMA[1,1]        0.05       30    29  
.   SIGMA[2,2]         1       29    39  
.   
. Total running time: 21.498 seconds
```



PopED: Near-optimal design

```
poped_db_practical <- create.poped.database(  
  poped_db_final,  
  xt = c(0.5, rep(24, 3), 32, 168)  
)
```

```
FIM_practical <- evaluate.fim(poped_db_practical)  
get_rse(FIM_practical, poped_db_practical)
```

.	CL	V	KA	d_CL	d_V	d_KA	SIGMA[1,1]	SIGMA[2,2]
.	10.15307	19.61044	22.79548	48.67831	97.63716	74.56113	26.52843	40.69749

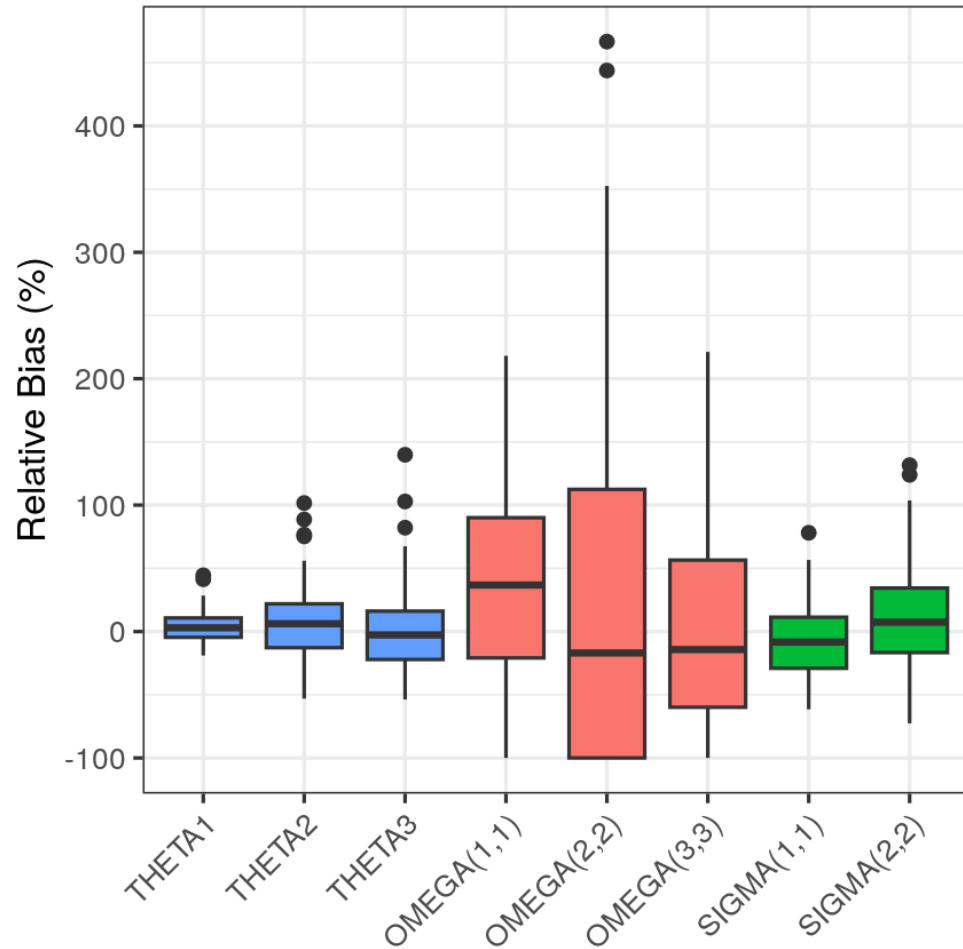


PopED: Sampling windows

```
plot_efficiency_of_windows(  
  poped_db_practical,  
  xt_plus = c(0.25, rep(0, 3), 2, 0),  
  xt_minus = c(0.25, rep(1, 3), 2, 4)  
)
```



SSE Results



param	poped_pct_rse	sim_pct_rse	sim_pct_bias
THETA1	10.2	12.2	4.2
THETA2	19.6	29.0	6.7
THETA3	22.8	32.0	0.4
OMEGA(1,1)	48.7	74.2	41.7
OMEGA(2,2)	97.6	132.8	25.9
OMEGA(3,3)	74.6	90.0	7.2
SIGMA(1,1)	26.5	28.6	-6.7
SIGMA(2,2)	40.7	41.3	11.9



The PFIM setup

PFIM requires types 3 of objects in order to define a `StatisticalModel`:

- `ModelEquations`, the structural model;
- `ModelParameter`, the parameter models (including IIV and IOV);
- `Response`, including the residual error model.

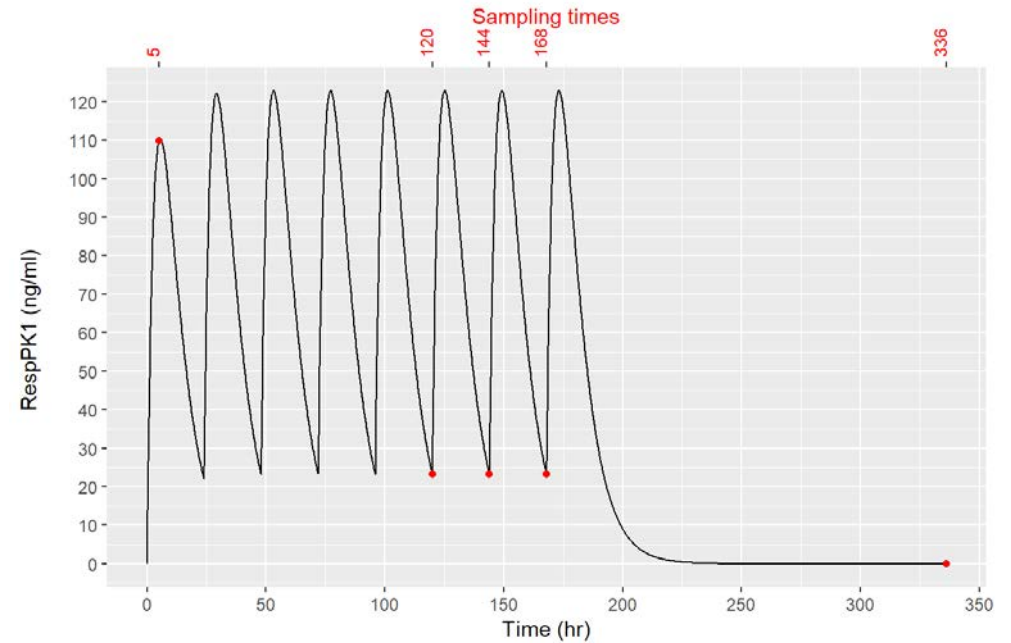
A `PFIMProject` object collects together these model objects with everything related to study design:

- `Arm` objects, which include `SamplingTimes` and `Administration` objects.



PFIM: Plot of initial design

```
evaluationPop <- EvaluatePopulationFIM(MyPro  
plotOptions <- list(  
  unitTime = c("hr"),  
  unitResponses = c("ng/ml")  
)  
  
plotResponse <- plotResponse(  
  evaluationPop,  
  plotOptions  
)  
  
print(plotResponse[[1]])
```



Design: design1 Arm: Bras test



P F I M: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)  
show(evaluationPop)
```

```
***** design1 *****  
  
  Arm_name Response tau Tinf                Time_dose  
1 Bras test  RespPK1  -    - 0, 24, 48, 72, 96, 120, 144, 168 10000, 10000, 10000, 10000, 10000
```



PFIM: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)
show(evaluationPop)
```

```
*****
```

```
Fisher information matrix
```

```
*****
```

```
*** Fixed effect
```

	μ_{Cl}	μ_{ka}	μ_V
μ_{Cl}	0.594311668	-0.8211656	-0.002425272
μ_{ka}	-0.821165609	564.9226170	-2.218057475
μ_V	-0.002425272	-2.2180575	0.008762577



PFIM: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)  
show(evaluationPop)
```

```
*****
```

```
Fisher information matrix
```

```
*****
```

```
*** Variance components
```

	ω^2_{Cl}	ω^2_{ka}	ω^2_V	$\sigma_{inter_RespPK1}$	$\sigma_{slope_RespPK1}$
ω^2_{Cl}	147.1693161	0.1756023	0.2450810	0.1067300	7.591955
ω^2_{ka}	0.1756023	51.9429628	128.1192438	0.2065110	21.923776
ω^2_V	0.2450810	128.1192438	319.9281155	0.4876588	51.107443
$\sigma_{inter_RespPK1}$	0.1067300	0.2065110	0.4876588	34.2620161	238.748328
$\sigma_{slope_RespPK1}$	7.5919547	21.9237756	51.1074432	238.7483285	5561.894327



PFIM: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)  
show(evaluationPop)
```

```
*****  
Correlation matrix  
*****
```

```
*** Fixed effect
```

	μ_{Cl}	μ_{ka}	μ_V
μ_{Cl}	1	1	1
μ_{ka}	1	1	1
μ_V	1	1	1



PFIM: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)
show(evaluationPop)
```

```
*****
```

```
Correlation matrix
```

```
*****
```

```
*** Variance components
```

	ω^2_{Cl}	ω^2_{ka}	ω^2_V	$\sigma_{inter_RespPK1}$	$\sigma_{slope_RespPK1}$
ω^2_{Cl}	1.000000000	-0.007756317	0.007627033	0.003569392	-0.008783515
ω^2_{ka}	-0.007756317	1.000000000	-0.993851261	0.013130503	-0.027650113
ω^2_V	0.007627033	-0.993851261	1.000000000	-0.010893393	0.022752548
$\sigma_{inter_RespPK1}$	0.003569392	0.013130503	-0.010893393	1.000000000	-0.547261481
$\sigma_{slope_RespPK1}$	-0.008783515	-0.027650113	0.022752548	-0.547261481	1.000000000



P F I M: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)
show(evaluationPop)
```

```
*****
Determinant, condition numbers and D-criterion
*****
```

	[,1]
Determinant	4.394250e+00
Cond number fixed effects	1.718267e+14
cond number variance components	1.017806e+04
D-criterion	1.203263e+00



PFIM: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)  
show(evaluationPop)
```

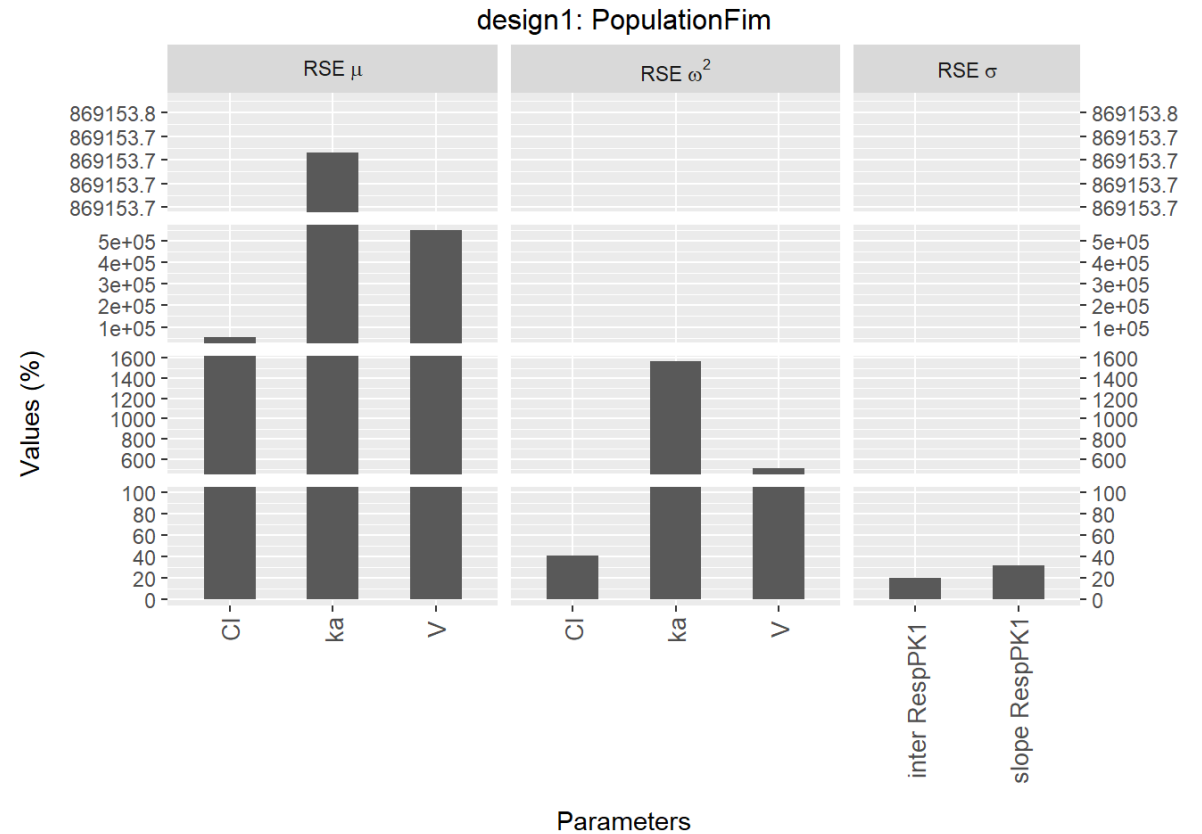
SE and RSE

	Value	SE	RSE (%)
μ_{Cl}	10.00	5.252748e+03	52527.47704
μ_{ka}	0.25	2.172884e+03	869153.72640
μ_V	100.00	5.514727e+05	551472.68815
ω^2_{Cl}	0.20	8.243712e-02	41.21856
ω^2_{ka}	0.08	1.254450e+00	1568.06291
ω^2_V	0.10	5.054008e-01	505.40077
$\sigma_{inter_RespPK1}$	1.00	2.041242e-01	20.41242
$\sigma_{slope_RespPK1}$	0.05	1.603788e-02	32.07577



PFIM: Plot RSE

```
plotRSE <- plotRSE(evaluationPop)  
print(plotRSE[[1]])
```



PFIM: D -optimal design: Starting from the original design

```
brasTestDesign1 <- addSampling(brasTestDesign1, SamplingTimes(  
  outcome = "RespPK1",  
  sample_time = c(5, c(120, 144, 168), 336)  
))  
  
samplingBoundsConstraint <- SamplingConstraint(  
  response = "RespPK",  
  continuousSamplingTimes = list(  
    c(0, 6),  
    c(119, 120),  
    c(143, 144),  
    c(167, 168),  
    c(264, 336)  
  )  
)
```



P F I M: *D*-optimal design: Starting from the original design

```
simplexOptimizer <- SimplexAlgorithm(  
  pct_initial_simplex_building = 20,  
  max_iteration = 5000,  
  tolerance = 1e-6  
)  
  
optimization_populationFIM <- OptimizeDesign(  
  MyProject,  
  simplexOptimizer,  
  PopulationFim()  
)  
  
show(optimization_populationFIM)
```

Optimal designs

4.98139743936754, 119.999815287607, 143.0000003895
167.999998299556, 271.44467773064

SE and RSE

	Value	SE	RSE (%)
μ_{Cl}	10.00	1.38874526	13.88745
μ_{ka}	0.25	0.22079467	88.31787
μ_V	100.00	56.80758006	56.80758
ω^2_{Cl}	0.20	0.08241346	41.20673
ω^2_{ka}	0.08	0.48045173	600.56467
ω^2_V	0.10	0.19874160	198.74160
$\sigma_{inter_RespPK1}$	1.00	0.20408418	20.40842
$\sigma_{slope_RespPK1}$	0.05	0.01571215	31.42429



PFIM: *D*-optimal design: Add sample after final (SS) dose

```
brasTestDesign1 <- addSampling(  
  brasTestDesign1,  
  SamplingTimes(  
    outcome = "RespPK1",  
    sample_time = c(5, c(120, 144, 168), 240, 336)  
  )  
)  
samplingBoundsConstraint <- SamplingConstraint(  
  response = "RespPK",  
  continuousSamplingTimes = list(  
    c(0, 6),  
    c(119, 120),  
    c(143, 144),  
    c(167, 168),  
    c(168, 336),  
    c(336, 336)  
  )  
)
```

Optimal designs

4.52576291331019, 119.000000458829, 143.0007915001
169.416690394867, 205.731885951118, 331.2662977881

SE and RSE

	Value	SE	RSE (%)
μ_{Cl}	10.00	1.29952262	12.99523
μ_{ka}	0.25	0.02873640	11.49456
μ_V	100.00	10.96078787	10.96079
ω^2_{Cl}	0.20	0.08235915	41.17958
ω^2_{ka}	0.08	0.05679475	70.99344
ω^2_V	0.10	0.05102656	51.02656
$\sigma_{inter_RespPK1}$	1.00	0.18241440	18.24144
$\sigma_{slope_RespPK1}$	0.05	0.01236994	24.73988



Example: ODE PK model



Study design and model

Fakinumab is being studied in humans for the first time.

- Single IV bolus doses of fakinumab: 0.03, 0.1, 0.3, 1, 3, and 10 mg.
- 6 subjects per dose group will be on active drug.
- Proposed samples: 1 and 4 hours post dose, and 1, 3, 7, 14, 21 days post dose.

Based on projections from animal PK data, we predict that a 2-compartment model with linear and nonlinear (Michaelis-Menten) clearance from the central compartment will describe the data.

CL	VMAX	KM	V1	Q	V2
0.5	20	1.2	2.5	10	4

We include log-normal IIV on CL, VMAX, and V1, and a proportional residual error.

om_CL	om_VMAX	om_V1	sigma_prop
0.2	0.2	0.1	0.15



Model in mrgsolve

```
.  
. Model file:  model_poped.mod  
. [ param ]  
. CL = 1, VMAX = 10, KM = 10, V1 = 8, Q = 10, V2 = 100  
.   
. [ cmt ] CENT PERIPH  
.   
. [ main ]  
. double ke  = CL/V1;  
. double k12 = Q/V1;  
. double k21 = Q/V2;  
.   
. [ ode ]  
. double CP = CENT/V1;  
.   
. dxdt_CENT = k21*PERIPH - k12*CENT - VMAX*CP/(KM + CP) - ke*CENT;  
. dxdt_PERIPH = k12*CENT - k21*PERIPH;  
.   
. [ capture ]  
. CP
```



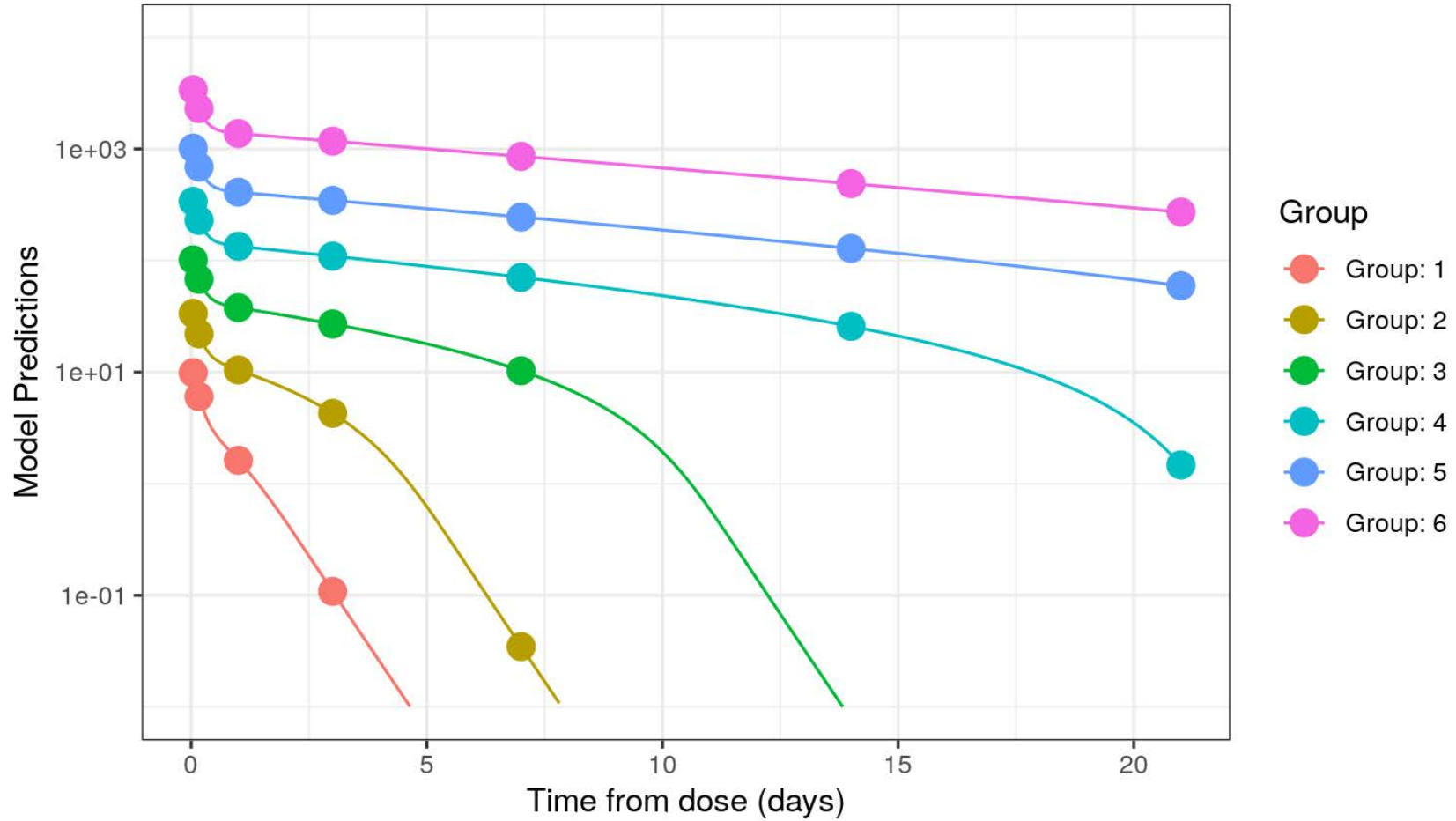
PopED: ff() for mrgsolve model

```
ff <- function(model_switch, xt, parameters, poped.db) {  
  obs_time <- as.numeric(xt)  
  dose_time <- 0  
  
  dose <- data.frame(  
    ID = 1,  
    amt = parameters[["DOSE"]]*1000,  
    cmt = 1,  
    evid = 1,  
    time = dose_time  
  )  
  obs <- data.frame(  
    ID = 1,  
    amt = 0,  
    cmt = 1,  
    evid = 0,  
    time = sort(obs_time)  
  )
```

```
  data <- arrange(bind_rows(dose,obs),time)  
  mod <- param(mod, parameters)  
  out <- mrgsim_q(mod,data,output="matrix")  
  out <- out[data$evid==0,"CP",drop=FALSE][match(obs_time,obs  
  return(list(y = out, poped.db = poped.db))  
}
```



PopED: Plot of initial design



PopED: Evaluate FIM

```
FIM_mrg <- evaluate.fim(poped_db_mrg)
get_rse(FIM_mrg, poped_db_mrg)
```

.	CL	VMAX	KM	V1	Q	V2	d_CL	d_VMAX	
.	10.874298	10.660146	7.935583	6.355286	8.894680	3.620177	38.989274	32.096484	31.66

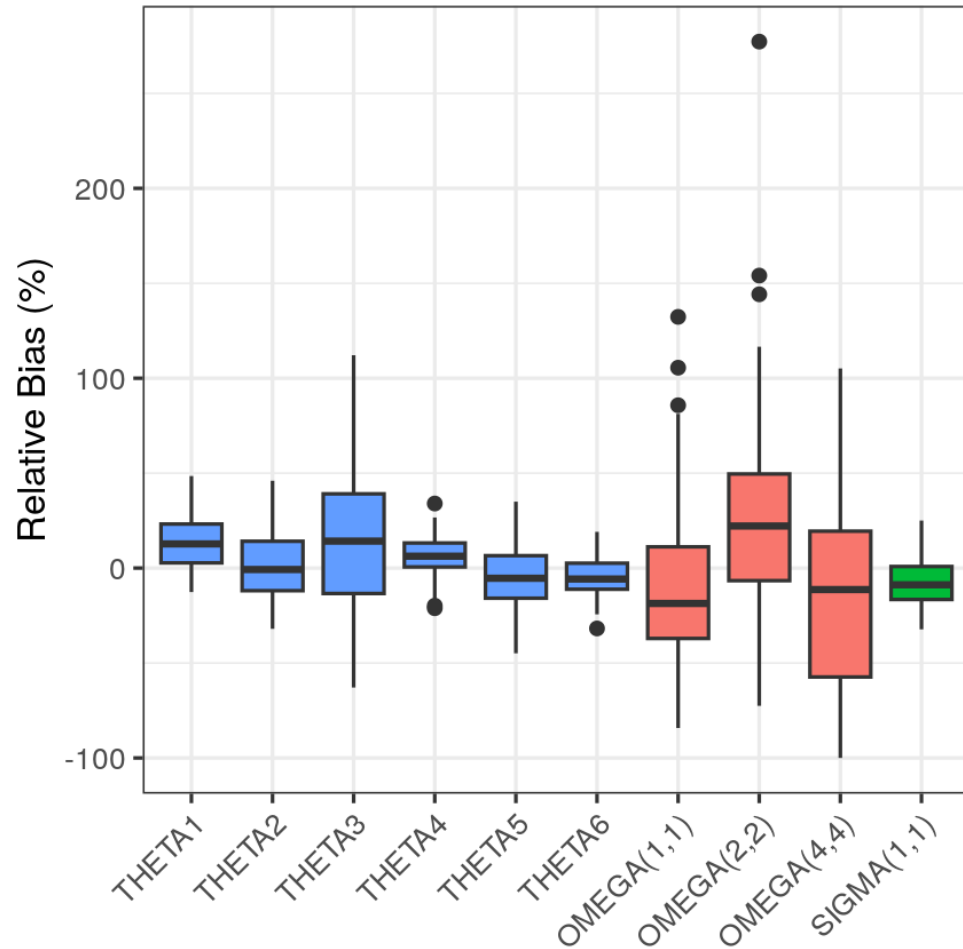


PopED: Sampling windows

```
plot_efficiency_of_windows(  
  poped_db_mrg,  
  xt_plus = c(rep(1/24, 2), rep(3/24, 5)),  
  xt_minus = c(rep(1/24, 2), rep(3/24, 5))  
)
```



SSE results



param	poped_pct_rse	sim_pct_rse	sim_pct_bias
THETA1	10.9	13.9	13.6
THETA2	10.7	17.1	1.8
THETA3	7.9	38.4	15.5
THETA4	6.4	10.4	6.6
THETA5	8.9	16.9	-5.8
THETA6	3.6	9.6	-4.4
OMEGA(1,1)	39.0	38.9	-11.9
OMEGA(2,2)	32.1	51.1	24.6
OMEGA(4,4)	31.7	52.3	-12.6
SIGMA(1,1)	10.8	12.3	-7.0



PFIM: ODE model

```
MyStatisticalModel <- StatisticalModel()
MyStatisticalModel <- setParametersOdeSolver(MyStatisticalModel, list(atol = 1e-16, rtol = 1e-6))

MyModelEquations <- ModelODEquations(
  list(
    "RespPK1" = expression(C1),
    "RespPK2" = expression(C2)
  ),
  list(
    "Deriv_C1" = expression((Q / V2) * C2 - (Q / V1) * C1 - VMAX * (C1 / V1) /
      (KM + (C1 / V1)) - (CL / V1) * C1),
    "Deriv_C2" = expression((Q / V1) * C1 - (Q / V2) * C2)
  )
)

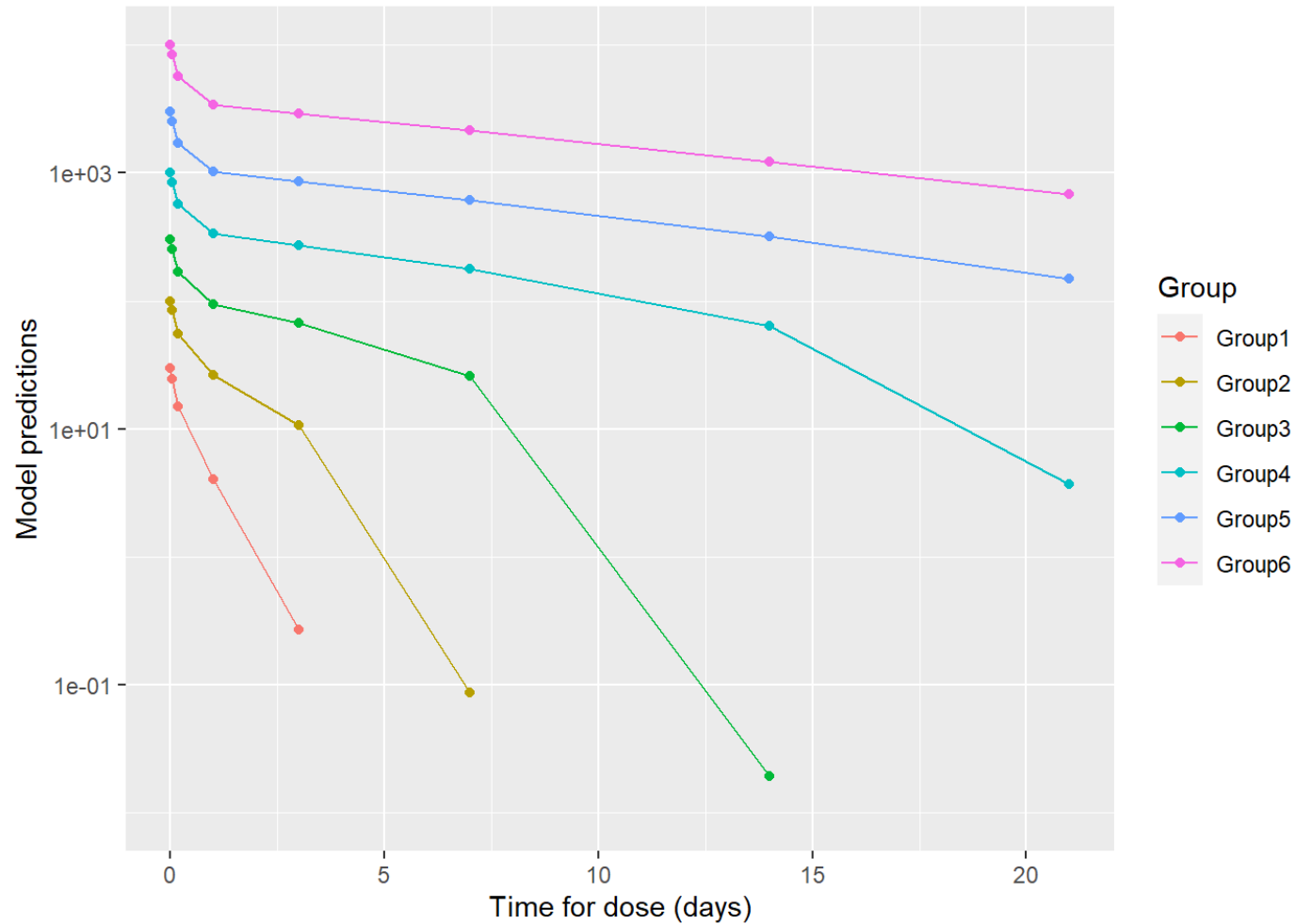
MyStatisticalModel <- defineModelEquations(MyStatisticalModel, MyModelEquations)

vC1 <- ModelVariable("C1/V1")
vC2 <- ModelVariable("C2")

MyStatisticalModel <- defineVariable(MyStatisticalModel, vC1)
MyStatisticalModel <- defineVariable(MyStatisticalModel, vC2)
```



P F I M: Plot of initial design



P F I M: Evaluate FIM

```
evaluationPop <- EvaluatePopulationFIM(MyProject)
show(evaluationPop)
```

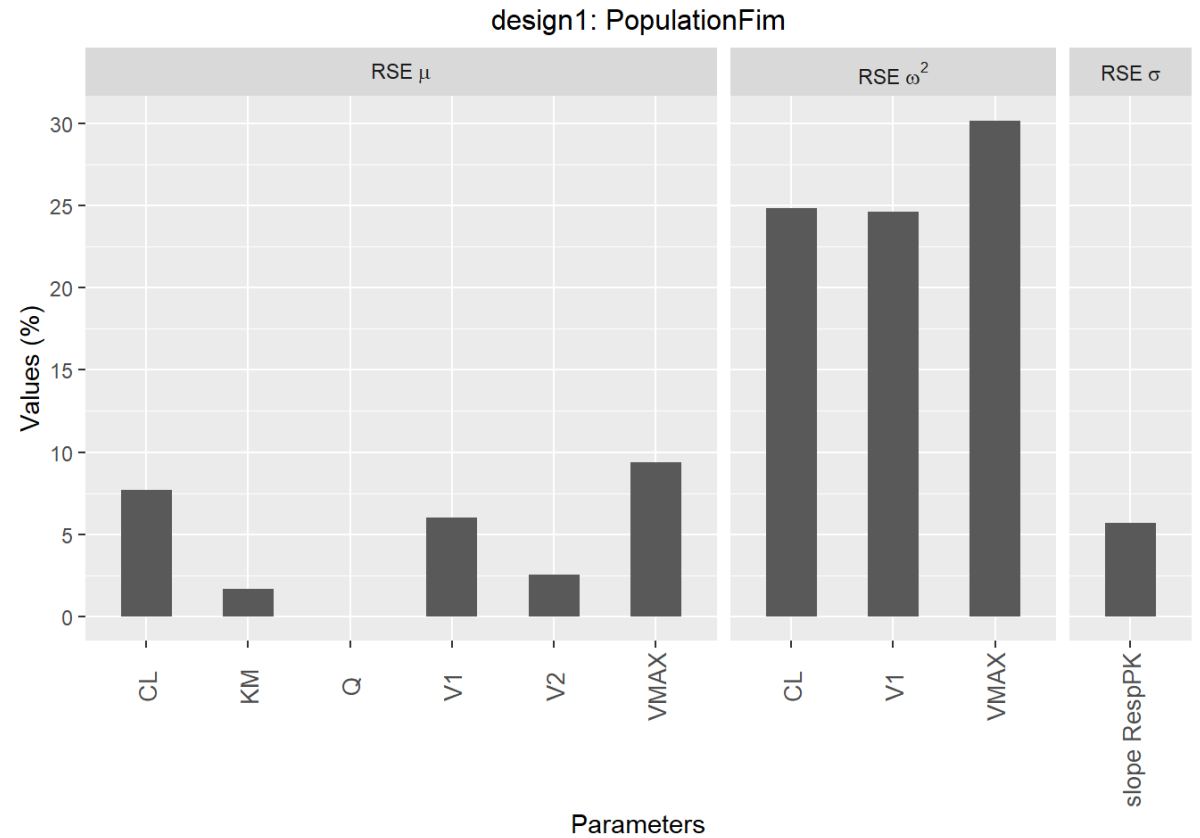
SE and RSE

	Value	SE	RSE (%)
μ_{CL}	0.50	0.0384775981	7.695519616
μ_{KM}	1.20	0.0196656516	1.638804302
μ_Q	10.00	0.0005779689	0.005779689
μ_{V1}	2.50	0.1498846589	5.995386356
μ_{V2}	4.00	0.1009500500	2.523751249
μ_{VMAX}	20.00	1.8703324957	9.351662478
ω^2_{CL}	0.20	0.0497454303	24.872715150
ω^2_{V1}	0.10	0.0246396407	24.639640672
ω^2_{VMAX}	0.20	0.0603291975	30.164598760
σ_{slope_RespPK}	0.15	0.0085116372	5.674424799



PFIM: Plot RSE

```
plotRSE <- plotRSE(evaluationPop)  
print(plotRSE[[1]])
```



Example: ODE PK/PD model



Study design & model

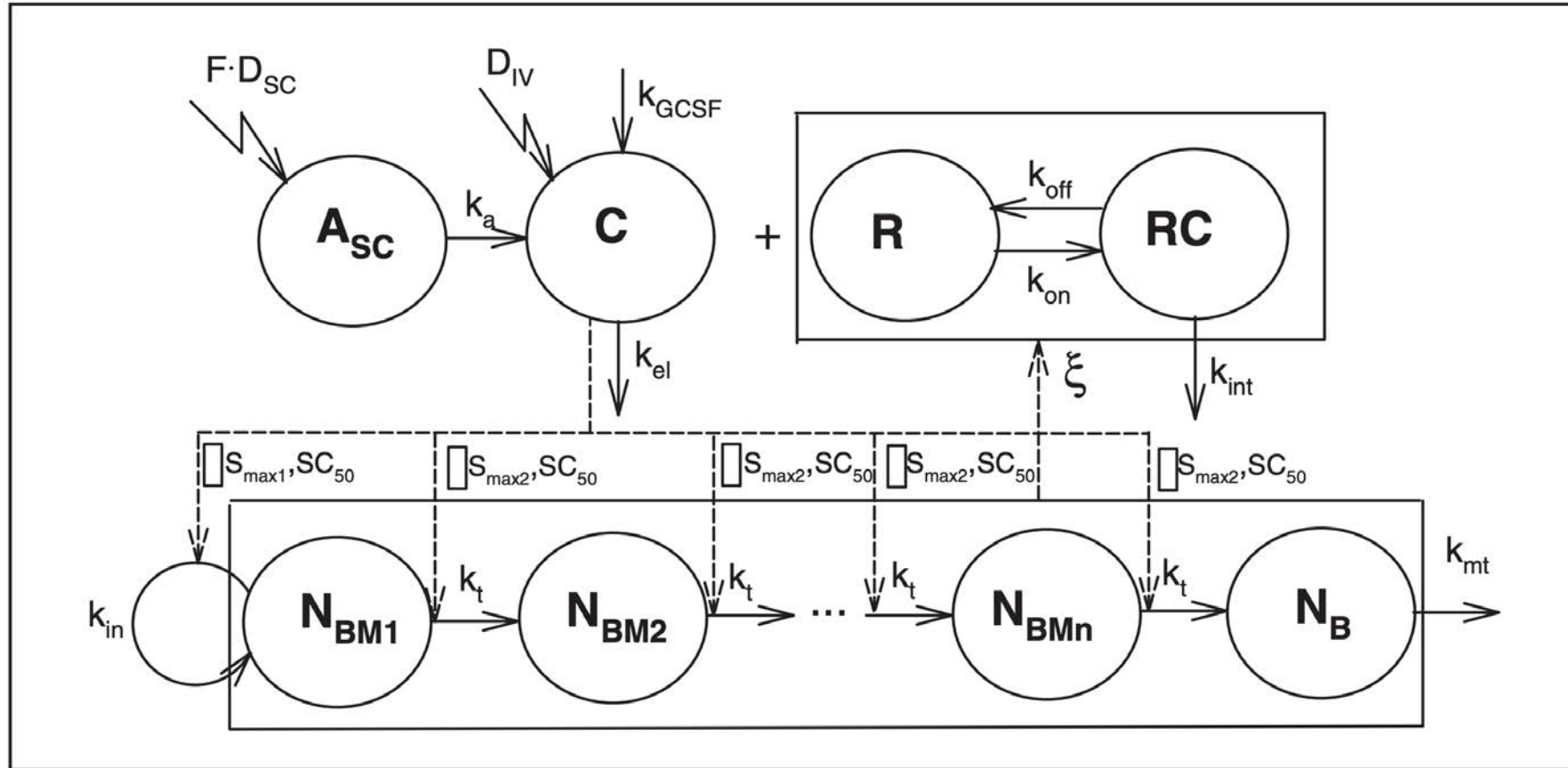
- QD SC doses of **filgrastim**: 1, 3, and 10 $\mu\text{g}/\text{kg}$.
- 10 subjects per dose group will be on active drug.
- Dense PK and ANC samples on days 1 and 7.

Population Modeling of Filgrastim PK-PD in Healthy Adults Following Intravenous and Subcutaneous Administrations

*Wojciech Krzyzanski, PhD, Pawel Wiczling, PhD, Phil Lowe, PhD, Etienne Pigeolet, PhD,
Martin Fink, PhD, Alexander Berghout, MD, and Sigrid Balsler, PhD*

- PK/PD model from DDMORE
<http://repository.ddmore.eu/model/DDMODEL00000077.6>
- NONMEM model translated to mrgsolve
<https://github.com/mrgsolve/depot/blob/master/vignette/gcsf.md>

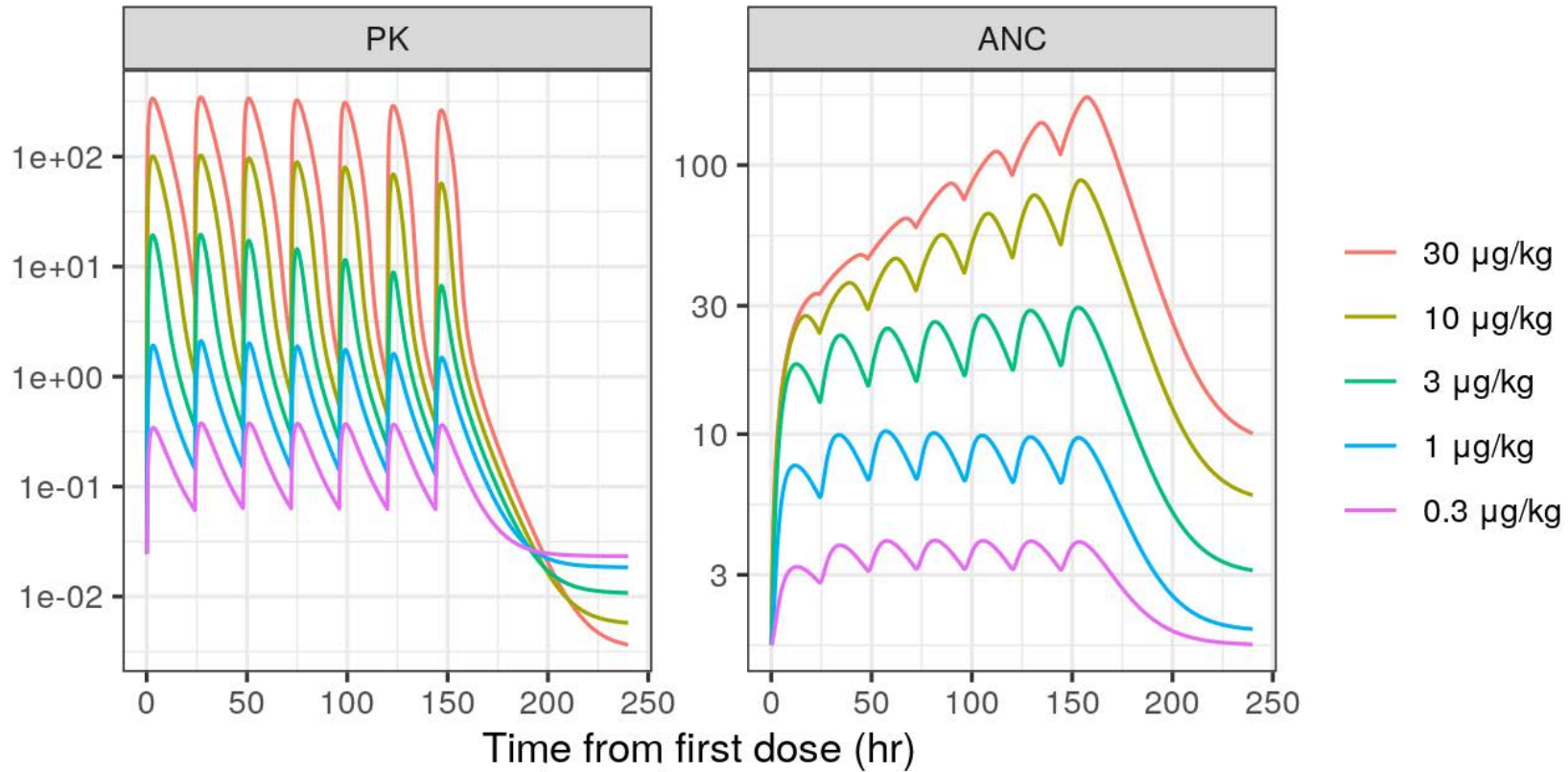
Filgrastim ANC model



Krzyzanski, Wiczling, Lowe et al. (2010)



PK and ANC simulations



Evaluate FIM for initial design

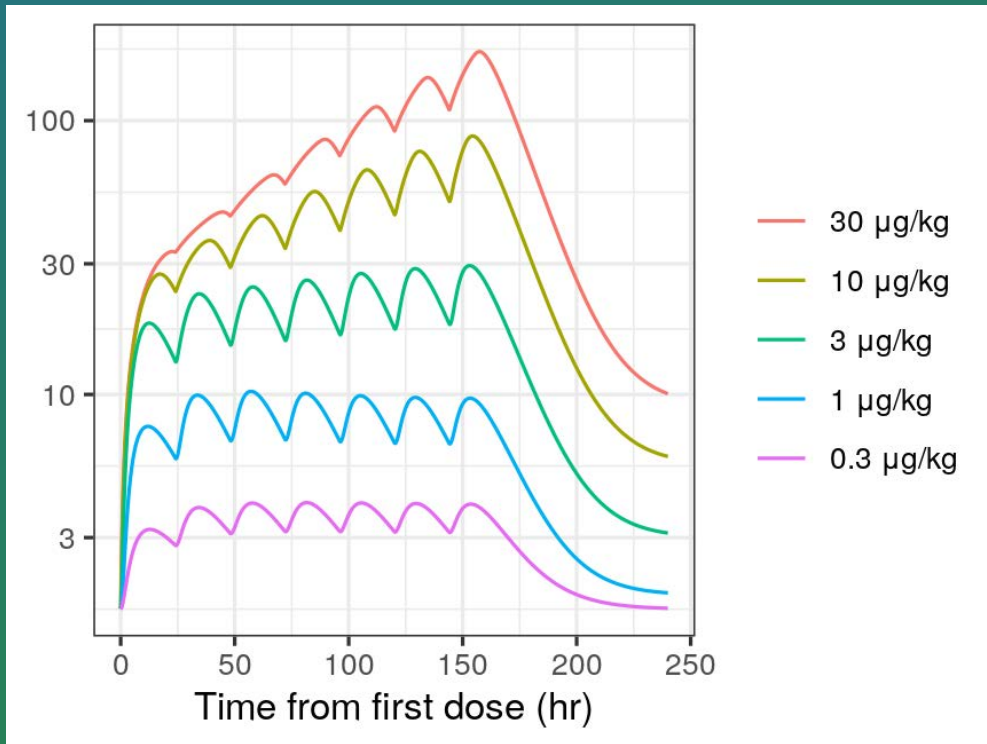
```
FIM_gcsf <- evaluate.fim(poped_db_gcsf)
get_rse(FIM_gcsf, poped_db_gcsf)
```

```
.          KA          KEL          VD          KD          KINT          KSI          KMT          KTT
.   9.332963  13.760898  12.690538  8.603883  3.911012  20.404806  3.418712  17.263477  4.00
.   d_SC1    d_SM1 SIGMA[1,1] SIGMA[3,3] SIGMA[4,4]
.  27.653544  26.362398  3.926952  6.505779  7.638103
```



Pop Quiz

Any better doses for estimating SC50 or Smax?



1. 1, 3, 10 µg/kg

2. 0.3, 1, 3 µg/kg

3. 3, 10, 30 µg/kg



Pop Quiz

Any better doses for estimating SC_{50} or S_{\max} ?

Parameter	Design 1	Design 2	Design 3
SC1	17.8	19.7	18.1
SM1	7.2	10.2	6.2
SM2	6.4	9.3	5.4



Wrap up



What did we learn today?

- Optimal design can be a useful, if imperfect, tool to explore and optimize study design options
- Always run confirmatory simulations
- Optimal design: it's not just for PK sampling any more!



What did we miss?

- Dealing with parameter uncertainty (e.g., ED -, $HC\ln D$ -optimality)
- Dealing with model structure uncertainty (e.g. T -optimality)
- Ignoring unimportant parameters (e.g. D_S -, G -optimality)
- Basically an alphabet of other criteria: A -, C -, G -, V -optimality, etc.
- Almost all options available in PopED and PFIM



Software resources

- PopED: <https://andrewhooker.github.io/PopED/>
- PFIM: <http://www.pfim.biostat.fr/>
- mrgsolve: <https://mrgsolve.github.io/>
- bbi:
<https://github.com/metrumresearchgroup/bbi>
 - bbr:
<https://metrumresearchgroup.github.io/bbr/>



References

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- Leroux, R., J. Seurat, H. Nagard, et al. (2022). *Design evaluation and optimisation in nonlinear mixed effects models with the R package PFIM*. Poster. Abstr 10183.
- Mentre, F., A. Mallet, and D. Baccar (1997). "Optimal design in random-effects regression models". In: *Biometrika* 84.2, pp. 429-442.
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- Retout, S., S. Duffull, and F. Mentre (2001). "Development and implementation of the population Fisher information matrix for the evaluation of population pharmacokinetic designs". In: *Comput. Methods Programs Biomed.* 65.2, pp. 141-151.
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Thank you

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Backup slides



Simulation



SSE: template.csv

```
$PROB RUN# run_num  
$INPUT ID TIME EVID MDV CMT AMT SS II DV WT  
$DATA data_fname IGNORE=@  
...  
$SIMULATION (run_num)  
$ESTIMATION METHOD=1 INTER PRINT=1 MSFO=./run_num.msf
```



SSE: Run the models with bbr

```
run_model <- function(.design_dir, .run_num) {  
  # Modify and write the control stream  
  ctl_template %>%  
    str_replace("run_num", as.character(.run_num)) %>%  
    str_replace("data_fname", paste0("../", .run_num, ".csv")) %>%  
    writeLines(file.path(.design_dir, paste0(.run_num, ".ctl")))  
  
  # Run the model  
  new_model(  
    .yaml_path = paste0(.run_num, ".yaml"),  
    .description = .run_num,  
    .directory = design_dir  
  ) %>%  
    submit_model()  
}
```



SSE: Collect the results with bbr

```
get_est <- function(.design_dir) {  
  est <- map_dfr(seq_len(n_rep), function(.run_num) {  
    mod <- read_model(paste0(.run_num, ".yaml"), .directory = .design_dir)  
    mod_sum <- try(model_summary(mod), silent = TRUE)  
    if (inherits(mod_sum, "try-error")) return(NULL)  
    mod_sum %>%  
      param_estimates() %>%  
      filter(fixed == 0) %>%  
      select(param = names, estimate) %>%  
      mutate(rep = .run_num)  
  }) %>%  
  left_join(true_vals) %>%  
  mutate(  
    param = factor(param, levels = unique(.[["param"]])),  
    pct_bias = (estimate - value) / abs(value) * 100  
  ) %>%  
  filter(abs(pct_bias) < 5000)  
  return(est)  
}
```



PopED setup



ff () : the structural model

```
ff <- function(model_switch, xt, parameters, poped.db)
  with(as.list(parameters),{

    CL <- CL*(WT/70)^(WT_CL)
    V <- V*(WT/70)^(WT_V)

    y_sd <- (DOSE * KA/(V * (KA - CL/V))) *
      (exp(-CL/V * xt) - exp(-KA * xt))

    y_ss <- (DOSE * KA/(V * (KA - CL/V))) *
      (exp(-CL/V * xt) / (1 - exp(-CL/V * TAU)) -
        exp(-KA * xt) / (1 - exp(-KA * TAU)))

    y <- xt
    y[model_switch == 1] <- y_sd[model_switch == 1]
    y[model_switch == 2] <- y_ss[model_switch == 2]

    return(list(y = y, poped.db = poped.db))
  })
}
```

PopED expects a function with the following arguments:

- `model_switch`: A vector of values identifying which model response should be computed for the corresponding `xt` value
- `xt`: A vector of independent variable values (often time).
- `parameters`: A named list of parameter values.
- `poped.db`: A PopED database.



fg(): the parameter model

```
fg <- function(x, a, bpop, b, bocc){  
  parameters = c(  
    CL      = bpop[1] * exp(b[1]),  
    V       = bpop[2] * exp(b[2]),  
    KA      = bpop[3] * exp(b[3]),  
    WT_CL   = bpop[4],  
    WT_V    = bpop[5],  
    DOSE    = a[1] * 1000,  
    TAU     = a[2],  
    WT      = a[3]  
  )  
  return(parameters)  
}
```

- x: A vector of discrete design variables (not used here).
- a: A vector of covariates.
- bpop: A vector of fixed effect parameters (i.e., THETAs).
- b: A vector of individual IIV random effects (i.e., ETAs).
- bocc: A vector of individual IOV random effects (i.e., ETAs) (not used here).

In this example, we include IIV on CL, V, and KA, and pass through dose, tau, and body weight as covariates.



feps () : the residual error model

```
feps <- function(model_switch, xt, parameters, epsi, p
  returnArgs <- do.call(
    poped.db$model$ff_pointer,
    list(model_switch, xt, parameters, poped.db)
  )
  y <- returnArgs[[1]]
  poped.db <- returnArgs[[2]]
  y = y * exp(epsi[, 1])
  return(list(y = y, poped.db = poped.db))
}
```

- `epsi`: A matrix of residual random effects (i.e. EPSs or ERRs).



create.poped.database()

```
poped_db <- create.poped.database(  
  ff_fun = ff,  
  fg_fun = fg,  
  fError_fun = feps.add.prop,  
  bpop = c(CL = 10, V = 100, KA = 0.25, WT_CL = 0.75, WT_V = 1),  
  notfixed_bpop = c(1, 1, 1, 0, 0),  
  d = c(CL = 0.08, V = 0.1, KA = 0.2),  
  sigma = c(0.05, 1),  
  m = 1,  
  groupsize = 12,  
  xt = c(5, c(rep(24, 3), 168)),  
  minxt = c(0, c(rep(23, 3), 96)),  
  maxxt = c(6, c(rep(24, 3), 168)),  
  model_switch = c(1, rep(2, 4)),  
  a = cbind(DOSE = 10, TAU = 24, WT = 32)  
)
```



create.poped.database()

```
poped_db <- create.poped.database(  
  ff_fun = ff,  
  fg_fun = fg,  
  fError_fun = feps.add.prop,  
  bpop = c(CL = 10, V = 100, KA = 0.25, WT_CL = 0.75,  
  notfixed_bpop = c(1, 1, 1, 0, 0),  
  d = c(CL = 0.08, V = 0.1, KA = 0.2),  
  sigma = c(0.05, 1),  
  m = 1,  
  groupsize = 12,  
  xt = c(5, c(rep(24, 3), 168)),  
  minxt = c(0, c(rep(23, 3), 96)),  
  maxxt = c(6, c(rep(24, 3), 168)),  
  model_switch = c(1, rep(2, 4)),  
  a = cbind(DOSE = 10, TAU = 24, WT = 32)  
)
```

- ff_fun, fg_fun, fError_fun:
Model functions



create.poped.database()

```
poped_db <- create.poped.database(  
  ff_fun = ff,  
  fg_fun = fg,  
  fError_fun = feps.add.prop,  
  bpop = c(CL = 10, V = 100, KA = 0.25, WT_CL = 0.75,  
  notfixed_bpop = c(1, 1, 1, 0, 0),  
  d = c(CL = 0.08, V = 0.1, KA = 0.2),  
  sigma = c(0.05, 1),  
  m = 1,  
  groupsize = 12,  
  xt = c(5, c(rep(24, 3), 168)),  
  minxt = c(0, c(rep(23, 3), 96)),  
  maxxt = c(6, c(rep(24, 3), 168)),  
  model_switch = c(1, rep(2, 4)),  
  a = cbind(DOSE = 10, TAU = 24, WT = 32)  
)
```

- `bpop`: our current best estimates of the fixed effect parameters (THETAs)
- `notfixed_bpop`: whether or not they're being estimated
- `d`: diagonal elements of the IIV covariance matrix (OMEGA)
- `sigma`: diagonal elements of the residual covariance matrix (SIGMA)



create.poped.database()

```
poped_db <- create.poped.database(  
  ff_fun = ff,  
  fg_fun = fg,  
  fError_fun = feps.add.prop,  
  bpop = c(CL = 10, V = 100, KA = 0.25, WT_CL = 0.75,  
  notfixed_bpop = c(1, 1, 1, 0, 0),  
  d = c(CL = 0.08, V = 0.1, KA = 0.2),  
  sigma = c(0.05, 1),  
  m = 1,  
  groupsize = 12,  
  xt = c(5, c(rep(24, 3), 168)),  
  minxt = c(0, c(rep(23, 3), 96)),  
  maxxt = c(6, c(rep(24, 3), 168)),  
  model_switch = c(1, rep(2, 4)),  
  a = cbind(DOSE = 10, TAU = 24, WT = 32)  
)
```

- m: number of groups
- groupsize: number of subjects in each group



create.poped.database()

```
poped_db <- create.poped.database(  
  ff_fun = ff,  
  fg_fun = fg,  
  fError_fun = feps.add.prop,  
  bpop = c(CL = 10, V = 100, KA = 0.25, WT_CL = 0.75,  
  notfixed_bpop = c(1, 1, 1, 0, 0),  
  d = c(CL = 0.08, V = 0.1, KA = 0.2),  
  sigma = c(0.05, 1),  
  m = 1,  
  groupsize = 12,  
  xt = c(5, c(rep(24, 3), 168)),  
  minxt = c(0, c(rep(23, 3), 96)),  
  maxxt = c(6, c(rep(24, 3), 168)),  
  model_switch = c(1, rep(2, 4)),  
  a = cbind(DOSE = 10, TAU = 24, WT = 32)  
)
```

- xt: initial sampling design
- minxt: lower bound
- maxxt: upper bound
- model_switch: associate sampling times with model



create.poped.database()

```
poped_db <- create.poped.database(  
  ff_fun = ff,  
  fg_fun = fg,  
  fError_fun = feps.add.prop,  
  bpop = c(CL = 10, V = 100, KA = 0.25, WT_CL = 0.75,  
  notfixed_bpop = c(1, 1, 1, 0, 0),  
  d = c(CL = 0.08, V = 0.1, KA = 0.2),  
  sigma = c(0.05, 1),  
  m = 1,  
  groupsize = 12,  
  xt = c(5, c(rep(24, 3), 168)),  
  minxt = c(0, c(rep(23, 3), 96)),  
  maxxt = c(6, c(rep(24, 3), 168)),  
  model_switch = c(1, rep(2, 4)),  
  a = cbind(DOSE = 10, TAU = 24, WT = 32)  
)
```

- a: covariates



